Molloy University DigitalCommons@Molloy

Faculty Works: BCES (1999-2023)

Biology, Chemistry, Earth & Environmental Science (BCES)

Winter 2014

Does Stress Make You Fat?

Jodi F. Evans Ph.D. *Molloy College*, jevans@molloy.edu

Thomas Rhodes

Michelle Pazienza

Catherine Nunez

Follow this and additional works at: https://digitalcommons.molloy.edu/bces_fac

Part of the Biology Commons, and the Chemistry Commons DigitalCommons@Molloy Feedback

Recommended Citation

Evans, Jodi F. Ph.D.; Rhodes, Thomas; Pazienza, Michelle; and Nunez, Catherine, "Does Stress Make You Fat?" (2014). *Faculty Works: BCES (1999-2023).* 19. https://digitalcommons.molloy.edu/bces_fac/19

This Abstract is brought to you for free and open access by the Biology, Chemistry, Earth & Environmental Science (BCES) at DigitalCommons@Molloy. It has been accepted for inclusion in Faculty Works: BCES (1999-2023) by an authorized administrator of DigitalCommons@Molloy. For permissions, please contact the author(s) at the email addresses listed above. If there are no email addresses listed or for more information, please contact tochtera@molloy.edu.



46th Annual MACUB Conference Bergen Community College Paramus, New Jersey October 26, 2013













The Metropolitan Association of College & University Biologists

Serving the Metropolitan New York Area for 47 Years

> MACUB 2013-2014 EXECUTIVE BOARD MEMBERS

PRESIDENT Prof. Gary Sarinsky Kingsborough Community College

> VilCE-PRESIDENT Dr. Kathleen Nolan Saint Francis College

TREASURER Dr. Gerhard Spory SUNY College at Farmingdale

CORRESPONDING SECRETARY Dr. Paul Russo Bloomfield College

RECORDING SECRETARY Dr. Margaret Carroll Medgar Evers College

MEMBERS-AT-LARGE Dr. Carol Biermann Kingsborough Community College Dr. Michael Palladino Monmouth University Dr. Dirk Vanderklein Montclair State University Dr. Donald Stearns Wagner College

2013 CONFERENCE CHAIR Robert Highley Bergen Community College

2012 CONFERENCE CHAIR Dr. Alan R. Schoenfeld Adelphi University

2011 CONFERENCE CHAIR Dr. Tin-Chun Chu Co-Chair Dr. Angela V. Klaus Seton Hall University

> IN VIVO EDITOR Dr. Edward Catapane Medgar Evers College

AWARDS CHAIR Dr. Anthony DePass Long Island University

ARCHIVIST Dr. Kumkum Prabhakar Nassau Community College

Instructions for Authors

IN VIVO is published three times yearly during the Fall, Winter, and Spring. Original research articles in the field of biology in addition to original articles of general interest to faculty and students may be submitted to the editor to be considered for publication. Manuscripts can be in the form of a) full length manuscripts. mini-reviews short b) C) or communications of particularly significant and timely Manuscripts will be evaluated by two information. reviewers.

Articles can be submitted electronically to *invivo*@*mec.cuny.edu* or mailed as a printed copy (preferably with a diskette that contains the file) to the Editorial Board at Medgar Evers College. All submissions should be formatted double spaced with 1 inch margins. The title of the article, the full names of each author, their academic affiliations and addresses, and the name of the person to whom correspondence should be sent must be given. As a rule, full length articles should include a brief abstract and be divided into the following sections: introduction, materials and methods, results, discussion, acknowledgments and references. Reviews and short communications can be arranged differently. References should be identified in the text by using numerical superscripts in consecutive order. In the reference section, references should be arranged in the order that they appeared in the text using the following format: last name, initials., year of publication. title of article, journal volume number: page numbers. (eg. - ¹Hassan, M. and V. Herbert, 2000. Colon Cancer. In Vivo 32: 3 - 8). For books the order should be last name, initial, year of publication, title of book in italics, publisher and city, and page number referred to. (eg. - Prosser, C.L., 1973. Comparative Animal Physiology, Saunders Co., Philadelphia, p 59.). Abbreviations and technical jargon should be avoided. Tables and figures should be submitted on separate pages with the desired locations in the text indicated in the margins.

IN VIVO Editorial Board

Editor:

Dr. Edward J. Catapane, Medgar Evers College

Associate Editors: Dr. Ann Brown, Dr. Margaret A. Carroll, Medgar Evers College

In This Issue:

MACUB 2013-2014 Executive Board	inside cover
Instruction for Authors	inside cover
MACUB 2013 Conference Poster Presentation Award Winners	32
MACUB 2013 Conference Poster Abstracts	37
MACUB 2013 Conference Member Presentations	81
Conference Highlights	84
2014 Benjamin Cummings/MACUB Student Research Grants	85
Affiliate Members	inside back cover

2014 Benjamin Cummings/MACUB Student Research Grants

Application is now open

see page 85

has anti-inflammatory activity (Nguyen A, personal communication) and we have data that suggest it is involved in cell signaling. MTE4a also produces a compound with antibiotic activity that has been partially purified. We describe here the various assays performed using MTE4a grown both in solid and liquid media to elucidate its potential to produce active secondary metabolites.

Does Stress Make You Fat? T. Rhodes, M. Pazienza, C. Nunez and J.F. Evans, Molloy College, Rockville Centre, New York, USA.

ACTH is a major hormone of the stress axis or hypothalamic-pituitary-adrenal (HPA) axis. It is derived from pro-opiomelanocortin (POMC) the precursor to the melanocortin family of peptides. POMC produces the biologically active melanocortin peptides via a series of enzymatic steps in a tissue-specific manner, yielding the melanocyte-stimulating hormones (MSHs). corticotrophin (ACTH) and β -endorphin. The melanocortin system plays an imperative role in energy expenditure, insulin release and insulin sensitivity. Bone marrow derived mesenchymal stem cells circulate in the blood stream and as progenitor cells have the potential to differentiate into many cell types such as osteoblasts, chondrocytes and adipocytes. Here we examine the effects of ACTH on the mouse D1 bonemarrow derived MSC. ACTH significantly increased lipid accumulation during the adipogenic differentiation of D1 cells in a concentration- dependent manner. ACTH also shifts the temporal pattern of D1 adipogenic differentiation to the left i.e. differentiation occurs earlier with ACTH treatment. No significant differences in protein expression of peroxisome proliferator-activated receptor gamma (PPAR-y2), a regulating transcription factor of adipogenesis were found. Therefore the effects of ACTH are suggested to be mediated by an alternative pathway. Overall the results indicate a connection between increased adipose deposition and the elevated circulating ACTH associated with stress.

Preliminary Characterization of the *XRR1* Gene in the Yeast *S. cerevisiae.* Vanessa Rivera¹ and Marci J. Swede, Long Island University, Post, Brookville, NY.

We have characterized a novel gene, *XRR1* (eXhibits Rapamycin Resistance), whose null mutant exhibits temperature sensitive resistance

to the anti-fungal drug rapamycin. The XRR1 gene product was shown via a systematic yeast two-hybrid analysis (Uetz et al, 2000) to physically interact with FKBP12 (FPR1p). The FKBP12 protein is responsible for binding rapamycin and causing cell-cycle arrest via the TOR signaling pathway. The XRR1 gene exhibits sequence homology to the A1pp (Macro) domain specific for binding ADP-ribose. This domain is found in the C -terminus of the mammalian macroH2A histone variant. Initial characterization of XRR1 null mutants indicates that they have no overall growth defect across a temperature range of 25°C to 37° C. Further characterization of the XRR1 gene suggests a role in the rapamycin resistance pathway. We have observed that the XRR1 null mutation results in an increase in growth in the presence of 100 ng/ml rapamycin at 37°C compared to growth at 30°C when exposed to the drug. The degree of resistant growth observed in the null mutant is greater than that observed by wildtype isogenic yeast under the same growth conditions. This drug resistance is not observed when growth at 30°C. We propose a model in which the XRR1 gene product is involved in stabilizing the FKBP12-rapamycin complex, which is responsible for cell cycle arrest in response to rapamycin treatment. The Swede lab gratefully acknowledges Dr. Hinnebusch at the NICHD for the gift of mutant strains. This research is supported by a grant from the Dextra Baldwin McGonagle Foundation in support of undergraduate research to MJ Swede and by a faculty research development grant from LIU, Post to MJ Swede.

The Effects of Resveratrol Compounds on the Motility and Proliferation of F10 Melanoma Cells. Christian Rivoira¹, Valery Morris² and Susan Rotenberg², ¹Queensborough Community College, Bayside, NY and ²Queens College, Flushing, NY.

Resveratrol is a phytochemical found in grapes and wine and has been reported to possess anti-carcinogenic effects. The effects of several cis and trans isomers of resveratrol were tested on highly metastatic mouse B16 F10 cells to see the effect on cell motility. Cells were seeded onto 96-well plates, treated with each compound and then run through a proliferation assay. F10 cells were also plated on 6-well plates and allowed to grow until 100% confluent for a wound healing assay. These samples were then scratched and treated with the compounds.